



# INLAND BEEEMAIL

Monthly newsletter of the Inland Empire Beekeepers Association

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Presidents  
Corner:

## President's Corner:

This month I may actually be at Friday's meeting. I guess I should show up at least once every four or five months.

The latest information I have on Travis Sammons is all good news. He was released from the hospital yesterday and allowed to go home. He will be staying in California for several months as he continues to heal up. He has been walking and talking and has the use of both his arms now. It will be slow but hopefully continuous improvement over the next several months. I'm sure Bill Watts will give us the up to date info at the meeting. In a separate article I will discuss ways to donate funds to pay hospital bills and how to get additional information at a web site set up for Travis.

The education committee conducted the first ever package installation demonstration. By all accounts, to include an Internet video, it was a great success. Once again, please welcome the new graduates. Please reinforce the concept that continuing education is a must for all beekeepers. Every year is different.

All the hives in this area are just exploding. My main "problem" is the bees plugging out the queen with nectar. I constantly have to move in new foundation to give the queen somewhere to lay. (The bees are drawing it out very fast, as in a very few days.) Obviously, this is a problem everyone should have. Lots of swarms. John Sammons has caught well over a dozen. The one real problem we have seen is chalk brood. Chalk brood is a "California thing". Hopefully the tytan in powdered sugar will fix it.

Enough rambling.  
See everyone next Friday.

## Inland Empire Beekeepers Association

### Agenda 5/9/08

#### Welcome!

I'm Back!

#### Reports:

The Secretary's Report – Linda  
The Treasurer's Report - Julie  
Fair Reports  
WSBA Report  
Four Corner Bee Reports

#### Old Business:

Education Committee Report – Harry Smits

#### New Business:

Pay your dues - no dues no beemail

Meeting Adjourned

#### CLASS:

\* Annual Smoker Safety Class

## ***IEBA - Meeting Minutes*** ***March - Julie Watts for Linda Carney,*** **Secretary**

Darren our Vice President suggested we consider purchasing Audio-Visual equipment. The current equipment being used through the WSU-Master Gardener is not always available. It is on a first come first serve basis. There has also been some technical difficulty with using their system.

He would like us to think of ways our club can make money for this endeavor. Some idea's suggested included, a Spring Garage Sale, have each club member contribute a certain amount of money (\$\$), Darren suggested possibly having an auction. An individual may have some unwanted equipment we could use in the Auction. Please continue to think of ideas.

We will also need to take a look at the cost and do some pricing comparisons. We have decided at this time there are enough people who have lap tops that it is not necessary at this time to purchase one.

### **North Bee Yard**

Per Bob we had 19 hives going into winter. Currently we now 7 alive.

### **Local 4 Corners Report**

Joe Jovanovich, his hives were doing great one month, then the next he had dead outs, he believes his was from *Nosema sp.*

Darren has lost 2 hives, 7 are currently questionable. The 2 dead outs were not previously treated.

### **WSBA Jerry's Report**

Eric Olson gave a crash course on *Nosema cerena*. They believe they now have a good protocol for Bee Keepers to follow. This should be finalized soon. The State appropriated \$20,000.00 for WSU for testing. King 5 news has been reporting many hives are crashing after coming out of the Orchards. Due to such high loss of Bees on the West side and not enough Bee's, people are scrambling for contracts.

New packages- Jerry indicated the new packages were to have been treated prior to being shipped w/ fumagillin. Jerry recommended to treat in a few weeks.

We had to cancel the invitation to teach the children at the Ag-Days at the fair grounds this year due to lack of people volunteering. This was also true for the request from the local libraries to have a local Bee Keeper come in and educate this summer. This is highly unfortunate because the children are the future.

Ted Swenson is currently working on grading tests. Jack Knox indicated the Liability insurance is due in May and Kelly is in charge of the North Idaho Fair.

A Swarm removal list is going around for anybody who would like to sign up. Please see Darren Mumau.

A Mentor list is also going around. Please sign up for this if you would like to help new bee keepers with questions. All help is appreciated. Both the Swarm Removal list as well as the Mentor list will be on-line next month.

### **Meeting adjourned.**

### **Treasure's report.**

With this meeting we had \$9182.58 in checking and \$1877.19 in the savings. This Figure does not include the cost for the new packages just received. This cost will be reflected in the next months report.



## **Travis Sammons Medical Benefit Fund**

A benefit fund has been established to help Travis Sammons and family pay for his recent medical expenses. The fund was set up at the U.S. Bank. Anyone wishing to donate can go to any US Bank and make a deposit into the 'Travis Sammons Medical Benefit Fund', account number 153560139609. All donations are greatly appreciated.

In addition, a web site has been set up with current information on Travis' medical condition. You may also donate using PayPal. The web address is <http://www.valleyadventist.org>. Click on Travis' picture to go to his page.

## Evaluating Your Colony & Your Queen

By: Jennifer Berry

### Check now for problems.

Courtesy of Bee Culture

When the calendar page is turned and the 1<sup>st</sup> week of March appears, we southerners realize that crunch time is upon us. There are only a few short weeks to get our colonies set and ready to go. Otherwise nectar will be left untouched and therefore unprocessed into honey. Hopefully we didn't spend the winter months goofing off but instead got plenty of work accomplished. Old equipment was repaired, ratty, black comb replaced, honey supers primed and ready for action and new, pristine apiary sites selected. If expanding operations then plenty of hammering, wiring, gluing, and painting were part of your Winter activities. If starting those first colonies then queens, packages or nucs have been ordered already. Whatever your plan of attack is I hope you are ready because the bees surely are.

Now that the equipment is in order let's see how the bees survived the winter. The first thing you will want to undertake this month is to inspect your colonies. Don't procrastinate! It is easy to put this off with other Spring-time chores breathing down your neck, but your bees may need you sooner than later. During the month of March there should be numerous opportunities to inspect your colonies. The earlier you finish this task the better. Assuming your colonies are ok by just observing bees flying in and out of the hive means nothing. On your first hive inspection of the year you really must open the hive and check each individual frame when the temperatures allow. Later in the year hive inspections don't need to be so thorough but you need a good idea how each colony is faring before the season begins.

So what are you looking for? Here are the basics. Is there a queen? How does the brood pattern look? Are there any signs of disease? How much honey and pollen is available and where is it located? How do the bees look? Are there signs of mites? And don't forget your notebook and pencil! Records on each individual hive are important information you will want to have, especially if something goes wrong down the road.

Let's begin the inspection with the most important issue; is there a queen and if so is she performing? If the colony is queenless then you may want to combine it with another colony, especially a weaker one. If you didn't order queens last year, getting one this Spring maybe almost impossible, especially this early. Now inspect the brood area. Brood patterns should be tight, with little to no skipped cells. The larvae should be pearly white. Discolored larvae could be a sign of disease or chilled brood. If you suspect a brood disease like AFB or EFB and are unsure how to diagnosis it, contact an experienced beekeeper or your county agent. The sooner a positive diagnosis can be reached the better. Chilled brood occurs when the brood nest expands too quickly for the bees to keep warm. The brood is exposed to cold temperatures and dies.

There should be an equal arrangement of eggs, larvae and capped brood. If the brood pattern is spotty, and the population low, at this point the best recommendation is to combine these colonies with others. There is no need allowing a colony to limp along if they aren't going to survive. These colonies are susceptible to disease, wax months, and robbing. By combining colonies you not only save the bees but the equipment as well. Just don't forget to kill the poor performing queen first before you combine. However, there are exceptions (doesn't there always seem to be exceptions when it comes to the world of beekeeping?). Some strains of bees will build up slower or faster depending on their genetics. Russians for instance are slower coming out the gate but will rapidly build up, catching or even surpassing your best colonies. You need to know the nature of your colony. In the past I've contemplated whether or not to combine certain weaker colonies. I didn't because the brood pattern was solid even though it was small. In a few cases the decision was a good one. They built up nicely and ended up making a substantial amount of honey. That is why good notes are an important asset. It helps you remember exactly what is happening in each colony.

Ok, the queen and brood appear to be in good shape, so how about the honey stores? Spring weather in the south can be very unpredictable. Last year we had one of the warmest Springs on record however that late two day Easter freeze in April wiped out not only the newly formed tender vegetation but colonies as well. Dramatic shifts in temperatures can separate the cluster from the food. Warm days the bees become active, then with sharp temperature drops the cluster can find itself separated from the honey stores. The bees may be only an

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inch away from the food but unable to retrieve it when temperatures plummet. The colony then starves before warmer temperatures arrive.

Even though the nectar flow is just around the corner don't count on it solely if honey stores are depleted. Colonies at this time are rapidly consuming food. Feeding each of those individual larvae takes a considerable amount of honey and pollen. They are nothing but little eating machines, made up primarily of a midgut and hindgut. And to think there are thousands of them per frame! So how much food is enough? This can be difficult to determine. However, the rule of thumb at our lab is too much is better than none. If our full size colonies are down to less than a half a super with no honey frames in the brood chamber, we feed. If we have surplus honey frames we add those, if not we use gallon baggies with syrup. Even though nectar flows may be only weeks away, inclement weather may keep the bees from flying and hence gathering nectar. Another thing to examine is the placement of the honey. As the cluster moves up into the honey supers during the Winter, honey is depleted in those areas. Move full frames of honey around the cluster. Frames of honey at the end of a super are worthless if the bees can't access them during cold spells.

And don't forget to check pollen stores. Here in the south the continued drought wreaked havoc on plant and animal life (as well as Atlanta's water supply). Little to no pollen was produced or collected. Mid Winter inspections of our colonies revealed absolutely no pollen. Not a single cell's worth. Therefore, add pollen patties now if your inspection reveals the same situation. There are numerous pollen substitute products available. Some are even pre-packaged into ready made patties which eliminate the hassle of having to mix it yourself. Pollen is the protein source needed for larval development. If there is little to no pollen, then brood production is reduced.

Even though the mite populations have decreased over the Winter months due to the decline in brood rearing, mites are still present. Examine the newly emerged bees to see if there are signs of deformed wings. If you see a considerable amount of deformed wings then treating should be on the horizon. However, we are nearing a nectar flow so

chemicals are out of the question. A non chemical approach to knock back mites is to dust adult bees with powdered sugar. The powdered sugar dislodges the mite from the adult bee. Used in conjunction with bottom screens or a sticky sheet, the mite is then removed from the hive. You will have to repeat this method several times in order to eliminate the mites emerging with workers and drones. The powdered sugar will not penetrate the wax cappings and therefore will not affect the reproductive or immature stage of the mite. After your inspection make sure you put the frames back in the order you removed them. You don't want to leave brood frames at the end of the hive because they'll be susceptible to colder temperatures.

Since we are only a few weeks away from the start of our nectar flow there is another issue we must consider. Overcrowded colonies are just itching to hit the trees once pollen and nectar start coming through the front door. If you want to make a substantial honey crop you need to discourage this natural, swarming tendency. One larger colony of 60,000 individuals has been shown to produce more honey than the honey combination of two smaller colonies with 30,000 individuals. Swarm prevention and control is important. There are many ways to accomplish this task but none are foolproof. Plus, once a colony has it in their mind to swarm, they will. The methods we choose is splitting, equalizing and cutting queen cells.

Colonies that are 'boiling over' with bees, (eight to 10 frames bees and brood) we split. We take four frames bees and brood (with eggs) and place them into a four to five frame nuc. If we have no queens available we allow the nuc to rear their own (which will take several weeks before the virgin queen will emerge). First of March in our area is a little early for queens to mate but by the end of March there should be ample drones and warm weather for mating. If there are weaker colonies in need of a frame or two of brood we take them from our stronger colonies and give it to them. Basically we rob Peter to feed Paul. Since we can't allow our breeder colonies to swarm we regularly cut queen cells. It's a painstakingly long process but until we have made our final selections we can't afford to lose a single queen.

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Retiring old, tired queens also helps to discourage swarming. Since my preference is to re-queen in the Fall the queen is only six months old when the swarm season hits. Another positive for Fall re-queening, there is no disruption to the colony just before the one and only nectar flow we experience. Our flow is short and sweet so we don't have time to mess around. There are Summer nectar flows to our north and south but this involves transporting hives. And finally, I already have a pretty good idea which queens are superior and which aren't so hot (because records are kept for each colony).

One more recommendation for swarm prevention, make sure the colony has plenty of room to expand. If you have empty, drawn deep frames drop those into the brood box. It gives the queen more cells to deposit eggs. Place these empty frames on the edge of the brood area. It's not a good idea to divide the cluster too early unless the colony is extremely strong. Removing old brood comb and replacing it with new wax foundation keeps the bees busy. Adding supers upstairs will also help ease congestion.

The last thing to discuss is site selection. This can be a difficult and time consuming chore but well worth the reward. If you have a few colonies and want those in your backyard, great. Just make sure they are facing south-east and aren't sitting in a low spot. Hive entrances facing the morning sun will warm up quicker thereby stimulating the colony to forage earlier (the early bird really does get the worm). Numerous nectar bearing plants only have nectar in the morning hours so you want your bees in the sky at first light. Other issues to be aware of when finding a site for your bees; Is there heavy agricultural activity in the area and if so what pesticides are being applied and when? Is there a clean source of water? Is it easily accessible, especially after it rains? How far is it? Are there wind breaks? Is it in a flood plain or water way? I have had to move my bees several times out of what seemed to be the perfect apiary site, but unfortunately no honey filled the supers. Then other sites which didn't look promising at all produced like mad. But don't get discouraged if you don't make honey the first year. You need to take into account weather conditions that year or the previous year. Give it a few seasons before abandoning a site.

Next month I'll discuss package installation since most packages are being produced and shipped at the end of March here in Georgia. But there is one final touch you can add to your empty hive bodies if packages are on

your calendar for delivery soon. Here in the south we are constantly battling small hive beetles. One thing we try to do with our newly constructed equipment is to caulk cracks, crevices and seams in the interior of the hive. They make perfect breeding grounds and hiding places for beetles. By sealing these areas the beetles are forced out in the open more often which in turn keeps the bees on their tails. Get those girls ready cause the flow is a coming.

See ya!

Jennifer Berry is the Research Coordinator at the University of Georgia Bee Lab.

## A Closer Look Queen Mating By: Clarence Collison

### Queens need multiple mates

Courtesy of Bee Culture

As we are learning more about the reproductive biology of the honey bee colony, we are discovering that there are biological advantages to queens mating with multiple drones. Virgin honey bee queens initiate mating very early in their lives, when they are approximately 1-2 weeks old. They take multiple flights and mate with numerous drones during each flight. Mating has profound and permanent effects on queen behavior, physiology, and resultant queen-worker interactions (Richard et al. 2007). Mating stimulates vitellogenesis (formation of yolk protein) and oocyte-maturation (egg formation) in the ovaries (Tanaka and Hartfelder 2004), which prompt the initiation of egg-laying. Furthermore, mating alters the pheromone profiles of queens, allowing them to regulate many different aspects of worker behavior and colony organization. While several other effects of mating on the physiology and behavior of queens are known, the effects of multiple-mating and semen quantity have not been extensively studied.

Richard et al. (2007) was able to demonstrate for the first time that insemination quantity significantly affects mandibular gland chemical profiles, queen-worker interactions, and brain gene expression. Because it is not possible to control the natural mating frequency of queens, instrumental insemination was used and queens

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inseminated with semen from either a single drone (single-drone inseminated, or SDI) or 10 drones (multi-drone inseminated, or MDI) were compared.

The chemical composition of the mandibular gland extracts of virgins, single- and multiple-mated queens were analyzed with gas chromatography, and a total of 27 compounds were found. There were significant differences in the chemical profiles between the three groups of queens. The greatest difference was between virgin queens and the two groups of mated queens, with 15 percent of the variation attributed to insemination quantity. Mated queens had significantly lower levels of 18 compounds relative to virgins, and higher levels of one compound. Multiple-mated queens had significantly lower levels of seven compounds than single-mated queens, and significantly higher levels of two compounds. Of the five original queen mandibular gland pheromone (QMP) components, the quantities of 9-ODA (9 oxo-(E)-2- decenoic acid), 9-HDA (9-hydroxy-(E)-2-decenoic acid) and HVA (4-hydroxy-3- methoxyphenylethanol) were all significantly lower in mated queens compared to virgins. Levels of 9-ODA and 9-HDA were also significantly lower in the multiple- than in the single-mated queens (Richard et al. 2007).

The relative proportions of the 27 individual compounds were compared to total gland quantity between virgin and mated queens and between single-mated and multiple-mated queens. The relative proportion of 13 compounds was significantly different between virgins and mated queens and 10 of these compounds were significantly higher in mated queen mandibular glands. There were fewer compounds with significantly different relative proportions between single- and multiple-mated queens. Two compounds (8-hydroxyoctanoic acid and unidentified compound 5) were present in significantly higher proportions in single-mated queens than in multiple-mated queens, while six compounds (4-hydroxybenzoic acid, unidentified compound 2, (E)-dec-2-enedioic acid, palmitic acid, alkane 2 and stearic acid) were significantly higher in MDI than in SDI queens mandibular glands. Of the QMP components, relative levels of 9-ODA were significantly lower in mated queens compared to virgins, while HOB (methyl p-hydroxybenzoate) levels were significantly higher. None of the QMP components differed between

SDI and MDI queens (Richard et al. 2007).

These changes in queen mandibular pheromone profiles occur immediately or shortly after insemination, but additional modifications occur over time and may be associated with age, egg laying, or both (Richard et al. 2007). One of the most measurable effects of queen pheromone is the induced retinue response, in which workers are attracted to the queen from a short distance, then lick and antennate her. Following insemination, the retinue response to SDI and MDI queens was monitored in observation hives, twice a day for five days. MDI queens attracted significantly more worker bees in their retinue than the SDI queens in natural colony conditions. Also, worker retinue responses to the mandibular gland extracts of virgin, SDI and MDI queens were tested in cages containing four-day-old bees. The retinue size was equivalent whether workers were exposed to virgin, SDI or MDI queen mandibular gland extracts. However, worker bees exposed to two different mandibular gland extracts at the same time preferred gland extracts from SDI and MDI to virgins and preferred MDI extracts to SDI extracts.

There are also profound neuro-endocrinal changes in the queen's brain after mating. Harano et al. (2005) measured the amount of dopamine and its metabolite (N-acetyldopamine) in the brain of six- and 12-day-old virgins and 12-day-old mated queens. Twelve-day-old mated queens showed significantly lower amounts of dopamine and its metabolite than both six- and 12-day-old virgin queens, whereas significant differences in the amounts of these chemicals were not detected between six- and 12-day-old virgin queens. These results are explained by down-regulation of both synthesis and secretion of brain dopamine after mating. It is speculated that higher amounts of brain dopamine in virgin queens might be involved in activation of ovarian follicles arrested in pre-vitellogenic stages, as well as regulation of their characteristic behaviors.

Brain expression levels were measured in single- and multiple-mated queens for a gene associated with phototaxis in worker honey bees (*Amfor*, the foraging gene) (Richard et al. 2007). Expression levels for this phototaxis gene were significantly altered by insemination quantity. *Amfor* expression

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levels in the queens' brains were significantly higher in single-mated queens than in multiple-mated queens. These differences are believed to be associated with changes in flight and phototaxis. Fully inseminated queens cease to be phototactic and no longer take mating flights. Previous studies have demonstrated that queens inseminated with lower quantities of semen (less than 8 µL) are more likely to continue to take mating flights (Woyke et al. 1995).

Clearly, mating number and insemination quantity have profound effects on queen physiology and behavior. Further research will be necessary to understand the mechanistic basis for these changes in queen behavior and physiology. The number and duration of mating flights, the physical act of insemination, the volume of ejaculate, quantity and viability of sperm, seminal proteins in the ejaculate and the genetic diversity of sperm and seminal proteins may all be important factors contributing to the changes in honey bee queen physiology, queen behavior, and social interactions in the colony. Both assays with live queens and cage studies with queen mandibular pheromone extracts demonstrate that workers can detect differences in pheromone profiles and will respond differently to virgins versus newly inseminated queens as well as singly versus multiply inseminated queens (Richard et al. 2007).

Since semen quantity affects the quality of the queen, beekeepers need to have queen mating occur in areas where there is a large population of sexually mature drones available. It takes numerous drones to fully inseminate a queen and there are clear direct benefits of a queen mating with multiple drones (Tarpy 2003). A fully filled spermatheca results in a longer egg-laying life span (Cobey 2003).

There are also many indirect benefits to multiple matings which may have equally important consequences for colony health and productivity. Drones carry different genes for a wide variety of traits. So when a queen produces worker offspring sired by different males, the workers vary from each other genetically (Tarpy 2003). The result of this increased genetic diversity is a variable and cosmopolitan worker population. Previous studies have demonstrated that there may be colony-level adaptive benefits for genetically diverse workers, i.e. disease resistance. Queens that mate with many drones will ensure that some of her workers (half on average) will inherit favorable alleles (one of a pair of genes for contrasting traits) from their father. A queen

that mates with only a single drone runs the risk of carrying alleles that are susceptible to a particular disease; queens that mate with multiple drones ensure that at least some of her workers will be resistant to the disease. Theoretically, any trait – anatomical, physiological, or behavioral – could be impacted by increased genetic diversity within a colony as a result of the queen mating with multiple drones. Beekeepers need to be sure that their queens are properly inseminated with a generous assortment of drone genotypes, so that the worker population is similarly variable.

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Clarence Collison is a Professor of Entomology and Head of the Department of Entomology and Plant Pathology at Mississippi State University, Mississippi State, MS.



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## HONEYBEE SWARM REMOVAL

NAME	PHONE	AREA
Bill & Julie Watts	509-448-1141 Cell 509-998-1261	Spokane
Will Olsen	509-482-7890	Spokane / Deer Park
Duane Hanna	208-659-7544	Kootenai County
Charles Gross	509-434-6249	Spokane / Lincoln CO
Joan Nolan	509-924-3652	Spokane Val- ley /  Spokane
Don Nilles	509-928-3616	Trentwood / Otis Orchards
Jim Lynn	208-623-5661	North Idaho
Peggy & Steven Abbott	509-928-4208	Mt. Spokane/ North side
Dean Cannon	509-744-9242	Spokane
Harry Smits	509-238-4118	North Spo- kane  Mead /  Chattaroy
Rick Sherman	509-928-3427	Spokane Val- ley
Jack Knoz	208-773-5452	Kootenai County
Dave Bearden	509-226-5231	Otis Orchards / Post Falls

**Letter To Gov. Gregoire Regarding Funding To Help Honeybees** – The Washington State Tree Fruit Coalition has sent a letter to Gov. Chris Gregoire in an effort to apprise her of the issues surrounding the disappearance of honeybees due to Colony Collapse Disorder (CCD). CCD has had devastating effects on hives sometimes killing up to 80 percent of the hive. In the letter the fruit industry is asking the governor to support research to identify the cause of CCD with an allocation of \$40,000 from her Emergency Fund. This amount would be added to a pledge of approximately \$70,000 that would create a diagnostic allocation at WSU. The overall funding would support analysis of affected hives and provide a methodology to treat the disease in order to keep hives viable.

## Beekeepers Fund Research

Two leading beekeepers in the Pacific Northwest, Eric Olson of Yakima and Tom Hamilton of Nampa, Idaho, have made donations to Washington State University as seed money for research to find out the cause of a phenomenon known as Colony Collapse Disorder, which has wiped out thousands of beehives through the region.

Noyes Apiaries in New Plymouth, Idaho; the Idaho Honey Association; and the Washington State Beekeepers' registration fund also have made contributions. With those donations and dedicated funds from the WSU Agricultural Research Center, researchers will spend nearly \$200,000 over the next two years.

The Washington State Beekeepers Association estimates that Washington's beekeepers have lost between 35 and 50 percent of their bees in recent years, according to a press release from WSU. Olson, who lost 4,000 hives worth approximately \$1.2 million this spring, said investing in the research and paying for any treatment that is found would be well worth the expense. "The most expensive thing I have is a dead beehive," he said.

## IEBA MENTORS

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John Pierce	509-242-2035	<a href="mailto:jpierce@jpe.net">jpierce@jpe.net</a>	South Hill Spokane
Dean Cannon	509-744-9242		Spokane

# Classified Ads

**Tate's Honey Farm** has all of your extracting and packaging needs as well as spring packages and queens. Woodenware for all your winter projects and spring needs. Shop hours are 8:30—2:00 every Saturday at E. 8900 Maringo, Millwood. Contact us at 509-924-6669 or online at [www.tateshoneyfarm.com](http://www.tateshoneyfarm.com)

## BEEBOXES BY LEE

Woodenware, standard or custom orders, IPM bottom boards, Hive top feeders, etc, select lumber. Order now to be ready for spring. Lee Berchtold  
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## NUC's - For Sale

Now taking orders for 08 Nucs 4.9mm and standard cell sizes available please call for prices. Contact Travis Sammons at 509-928-4326 / 509-991-3758

## Miller's Homestead

### Jim and Jenine Miller

Cheney, WA 1-509-299-9085  
14606 Stangland Rd., Cheney. Look at our web site for prices on all available items.  
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## NUC's For Sale

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open M-S 9-5:30

### East Farms Feed

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Otis Orchards, Wash. 99027  
509.928.3616

## IEBA Contacts

### President

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### IEBA Website

[www.inlandbeemail.com](http://www.inlandbeemail.com)

### WSBA Website

[www.wasba.org](http://www.wasba.org)

## Hive Care

# May:

# MAY

**The Bees.** Now the activity really starts hopping. The nectar and pollen should begin to come into the hive and may get heavy at times. The queen will be reaching her greatest rate of egg laying. The hive should be bursting with activity.

**The Beekeeper.** You can remove your Apistan strips (if they have been in the hive for 45 days). Also remove the menthol if you were using that for Tracheal Mite control. Add a queen excluder if you choose to use them, and place honey supers on top of the top deep. Watch out for swarming.

Inspect the hive weekly. Attend bee club meetings and workshops. And, if you are like rest of us, keep making up those frames to get ready in time for the honey flow.

If you have hives really busting at the seams, consider making up some splits to make up for any winter losses. An extra nuc or two always comes in handy.

-adapted from  
[www.backyardbeekeepers.com](http://www.backyardbeekeepers.com)





**Inland Empire  
Beekeepers  
Association**

**Next Meeting:  
Friday May 9th  
6:30 Meeting**

**T**he Inland Empire Beekeepers Association (IEBA) meets the 2nd Friday of every month at the Spokane County Ag Extension office by the County Fairgrounds, at 222 N. Havana. The association is affiliated with the Washington State Beekeepers Association (WSBA). IEBA membership dues are \$5.00 for an individual or \$10.00 for the entire family. This includes your receiving the *Inland Beemail*, which is published by the association every month.

## *INLAND BEEMAIL*

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### IEBA

#### Birthdays & Anniversaries

##### MAY BIRTHDAYS

- 13 -- Brian Spock
- 19 -- Scot Ingles
- 30 -- Lois Bremner

##### MAY ANNIVERSARIES

- 9 -- Roger & Linda Carney
- 17 -- Art and Clara Ross
- 18 -- Dave & Pam Zack

The Birthday and Anniversary lists need to be updated. If you are an IEBA member, and want to be included please send your name, along with the month/date of your birth. For the anniversary announcements please send the month/date and include your spouse's name. Send to: [joan1422@msn.com](mailto:joan1422@msn.com).

### *Beekeeping Calendar - Bob Arnold*

#### May

Check for AFB. Boomer colonies (2 or 3 boxes full of bees and brood) must be split/reduced or they will swarm. Move box with queen and open brood to a new stand 6 feet or so away. Introduce a mated queen immediately into the box on the original stand. Reduce colony strength, on those not split, by moving frames of brood and bees to weaker colonies. Replace queens that exhibit poor laying performance or those going into their second honey flow year. Feed pollen patties if insufficient pollen is available. Check weekly for swarm cells. Cut swarm cells only if there is evidence of a good laying queen (lots of eggs). Split colonies with swarm cells that are determined to swarm moving the box with the queen to new stand and leave cells in original stand. Keep switching brood boxes as top box gets full of brood. Keep checking new queens and package queens every 5 days for good egg laying. Replace new queen that is not laying properly immediately in the presence of nectar or syrup. Put 1:1 sugar syrup on during nectar dearth periods. First evidence of whitening comb put on new foundation in the top box. Add supers only if there is a substantial prolonged nectar flow. Start any swarms caught on foundation. Keep vegetation down around colonies.